

Figure S3. Testing the ability of electron transport chain inhibitors and a proton ionophore to increase *S. aureus* biofilm sensitivity to chloroxylenol. (**A to D**) Biofilm disruption assays on plastic were performed with *S. aureus* (Sa) Newman, chloroxylenol (Chlor) at 100 μg/ml, and the specified concentrations of 3-nitropropionic acid (3-NP) (**A**), Antimycin A (**B**), Sodium azide

(azide) (C), or oligomycin (Oligo) (D). Biofilms were grown for 6 hours, exposed to the above treatments for 18 hours, and S. aureus biofilm CFU were determined. (E) Biofilm disruption assays on plastic were performed with S. aureus (Sa) JE2 parental strain or the specified transposon mutant, and chloroxylenol (Chlor) at 100 µg/ml. Each column displays the average from at least two biological replicates, each with three technical replicates. Error bars indicate SD. ns, not significant; *, P < 0.05, ***, P < 0.001, by ordinary one-way ANOVA and Tukey's multiple comparison post-test. (F) Membrane potential was measured using the fluorescent dye DiOC₂ following exposure to carbonyl cyanide 3-chlorophenylhydrazone (CCCP) at 5 μM, HQNO at 100 μg/ml, Antimycin A at 100 μg/ml, or P. aeruginosa PA14 wild-type or ΔpqsLpvdApchE deletion mutant supernatant for 24 h. DiOC₂ fluorescence was measured at 680 nm following excitation at 485 nm. Results are reported as Fluorescence / OD₆₀₀. Each column displays the average from two biological replicates, each with three technical replicates. (G) Biofilm disruption assays on plastic were performed with S. aureus (Sa) Newman, chloroxylenol (Chlor) at 100 µg/ml, and CCCP at 5 µM or 25 µM. Biofilms were grown for 6 hours, exposed to the above treatments for 18 hours, and S. aureus biofilm CFU were determined. Each column displays the average from at least two biological replicates, each with three technical replicates. Error bars indicate SD. ns, not significant; *, P < 0.05, by ordinary one-way ANOVA and Tukey's multiple comparison post-test.